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Assessment of A Few Non Enzymatic Anti Oxidants in Selected Fruits

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ABSTRACT

The therapeutic benefits of the plant are often attributed to their antioxidant properties. The fruits and vegetables play an important role in providing antioxidants to the body. Antioxidants are needed to protect against free radicals that is produced in the body. Free radicals Free radicals damages the cells and play an important role in aging process and disease progression. The free radicals attack and damage the healthy cells and their structure and function are lost. This cell damage contributes to aging and degenerative diseases. The anti oxidant protection system is helpful in protecting the cells from reactive oxygen species. Plant derived materials are increasingly used for antioxidant activity. Plant metabolites function as protectants of cells. Antioxidants protect the body against oxidative stress by neutralizing free radicals. Plants contain rich amount of polyphenols which are very potent natural antioxidants. The present study was designed to evaluate the relative contribution of different polyphenols such as total phenolics, flavonoids and flavonol contents and their antioxidants activities. Now a days it is a trend to search for natural products to replace the synthetic ones. The present work was carried out with an objective to assess the non enzymatic antioxidants in the selected fruits(Ficus carica, Emblica officinalis, Cephalandra indica and Terminalia chebula). Among different plants studied Emblica officinalis fruits was found to contain more ascorbic acid, reduced glutathione, polyphenol, carotenoides and lycopene. The C.indica had least non enzymatic antioxidants. T. chebula extract had maximum α – tocopherol.

KEY WORDS: Antioxidants, F. carica, E. officinalis, C. indica and T. chebula

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INTRODUCTION

Globally a positive trend has blossomed in favour of traditional and integrative health sciences both in research and practices. Medicinal plants form a large group of important flora that are used for their therapeutic values. Antioxidants are important in the prevention of human diseases. Naturally occurring antioxidants in leafy vegetables and seeds such as ascorbic acid vitamin E and phenolic compounds possess the ability to reduce the oxidative damage associated with many diseases. Much of the protective effect of fruits and vegetables has been attributed to phytochemicals, which are the nonnutrient plant compounds such as the carotenoids, flavonoids, isoflavonoids, and phenolic acids. Thousands of phytochemicals have been identified in foods, yet there are still many that have not been identified. Different phytochemicals have been found to possess a range of activities, which may help in protecting against chronic disease. The phytochemicals may inhibit cancer cell proliferation, regulate inflammatory and immune response, and protect against lipid oxidation^{1,2} . A major role of the phytochemicals is protection against oxidation. We live in a highly oxidative environment, and many processes involved in metabolism may result in the production of more oxidants. Humans, and all animals, have complex antioxidant defense systems, but they are not perfect and oxidative damage will occur. Both cardiovascular disease and cancer are thought to be particularly the results of oxidative stress, which can lead to damage of the larger biomolecules, such as DNA, lipids, and proteins. It has been estimated that there are 10,000 oxidative hits to DNA per cell per day in humans³.

Ficus carica is a huge tree of the family Moraceae. The different parts of the tree namely bark, root, leaves , fruit and latex are used as remedy for various illnesses. It produces unique fruit which is an inverted flower. The species of Ficus are rich in polyphenolic compounds and flavanoids. These compounds have strong antioxidant properties that help in prevention and therapy of various oxidative stress related diseases⁴. The fruits and leaves are used as laxative, stimulant, antitussive, emollient, emmenagogue and resolvent^{5,6}.

Emblica officinalis Gaertn., known as amla, a member of Euphorbiaceae is extensively found all over India, as well as in Srilanka, Malaya, China, Pakisthan and Bangladesh. The fruits of the plant have been used in Ayurveda as a potent Rasayana which is used to promote health and longevity by increasing defense against disease. *Emblica officinalis* fruit extract has shown protective action against alcohol induced oxidative stress to the cells as evidenced by the lowered plasma transaminases, ALP, LDH and GT enzyme activities and elevated levels of the enzymic and non-enzymic antioxidants. In addition lowered cholesterol level and elevated HDL level demonstrate that *Emblica officinalis* fruit extract offers protection against cardiovascular risk. The active tannoid principles, poly phenolic

compounds and vitamin C present in AEEO could be contributed for the above mechanism⁷.Indian gooseberry (*Emblica officinalis* Gaertn.) (Euphorbiaceae) has a distinguished history in Ayurveda medicine and has ascribed a number of medicinal properties and as a dietary supplement, its use is increasing in Western countries. It is thought that its beneficial properties are a function of its antioxidant potency.

Cephalandra indica of the family Cucurbitaceae is commonly called as little gourd. It grows abundantly all over India, Tropical Africa, Australia, Fiji and throughout oriental countries. The plant is used extensively in Indian traditional system of medicine like Ayurveda and Unani⁸. The various parts of the plant are used by indigenous people to get relief from diabetes mellitus. The leaves are found to depress the activity of glucose 6 phosphatase and also has antioxidant activity⁹. *Cephalandra indica* roots extracted with hexane , chloroform, methanol, ethanol and water exhibited strong reducing power and total antioxidant capacity¹⁰.

Terminalia chebula belonging to the family combretaceae is an indigenous drug which forms a composition of various herbal formulations. It is reported to be antioxidant, hepato protective, antimicrobial^{11,12}, adaptogenic and anti-inflammatory¹³. Qercetin is an important component of T.chebula¹⁴.

Reactive oxygen species (ROS) are produced during metabolic and physiological processes. As a result of those, harmful oxidative reactions may oxidative/antioxidative balance shifts towards the oxidative status. Consequently, oxidative stress develops^{15,16}. Natural products such as herbs, fruits and vegetables become popular in recent years due to public awareness and increasing interest among consumers and scientific community¹⁷. Natural products which contain antioxidant properties such as phenolics, include flavonoids and phenolic acids ¹⁸, carotenoids and vitamins¹⁹. Epidemiological evidence has been provided that constituents in natural products show many biological and pharmalogical activities, including antioxidative, anti imflammatory and antiviral effects²⁰.Flavones, flavonols and proanthocyanidins are well known specific compounds associated with antioxidant activity in plants²¹. Phenolics can be classified into two groups, polyphenols and simple phenols which contain phenolic acids²². Most of the antioxidant properties in plants are also due to polyphenol, phenolic acid, flavonoid and vitamin C. The plants with high antioxidant activities also have high total phenolic and flavonoid content. Oxidative stress is caused by an imbalance between preoxidants and antioxidants. Antioxidants are radical scavengers which protect the human body against free radicals that may cause pathological conditions²³. Awareness of the importance of natural heritage and biodiversity is also growing. India is a gold mine of treasures with traditional and practical knowledge

of herbal medicines^{24,25}. The present work was carried out with an objective to assess the non enzymatic antioxidants in the selected fruits (*F. carica, E. officinalis, C. indica* and *T. chebula*).

METHODOLOGY

To estimate the ascorbic acid the sample was homogenised in 4% TCA and harvested, the supernatants were treated with a pinch of activated charcoal and incubated at 37[°] C for 15 minutes. Reaction mixture was centrifuged at 6000 rpm for 10 minutes to remove the charcoal residue. Supernatant was used to quantify vitamin C^{26} . For α –tocopherol²⁷ the sample was mixed slowly with 0.1 N sulphuric acid and incubated at room temperature for overnight. The reaction mixture was filtered through Whatman No 1 filter paper and filtrate is used for estimation. For reduced glutathione²⁸ equal volumes of tissue homogenate and 20% trichloroacetic acid were mixed. The precipitated fraction was centrifuged and to 0.025ml of supernatant 2ml of 0.6mM 5.5'- dithiobis(2-nitro benzoic acid) reagent was added. The final volume was made upto 3ml with phosphate buffer(0.2 M,pH 8.0). The colour developed was read at 412nm against blank. The polyphenol was estimated by Folin ciocalteu²⁹ procedure. The samples were prepared and 200 1 was introduced into test tubes.One ml of Folin ciocalteu reagent and 0.8 ml of sodium carbonate(7.5%) were added. The tubes were mixed and allowed to stand for 30 minutes. Absorption at 765 nm was measured. The total carotenoides³⁰ and lycopene³¹ in the sample are extracted in petroleum ether. The total carotenoids are estimated in UV/visible spectrophotometer at 450nm and same extract was read at 503 nm for lycopene. The results obtained were subjected to statistical analysis of one way ANOVA.

RESULTS AND DISCUSSION

It is evident from Table 1 that the highest values of ascorbic acid (1.241 mg/g), reduced glutathione (0.013 mg/g), polyphenol (0.129 mg/g), carotenoides (0.677 mg/g) and lycopene (0.019 mg/g) were observed in the fruit extract of *E.officinalis*. Fruit extract of *C.indica* was found to be the poorest source of the non-enzymatic antioxidants namely ascorbic acid (0.017 mg/g), reduced glutathione (0.006 mg/g), polyphenol (0.013 mg/g), carotenoides (0.020 mg/g) and lycopene (0.007 mg/g). Maximum amount of α – tocopherol (0.545 mg/g) was present in the extract of *T.chebula* and minimum (0.139 mg/g) in the extract of *E.officinalis*.

| S.No | Plant Screened | Non Enzymatic Anti Oxidants In The Plant Samples(mg/g) | | | | | |
|------|-------------------|--|-------------------|---------------------|------------|--------------|----------|
| | | Ascorbic acid | α – tocopherol | Reduced glutathione | polyphenol | carotenoides | Lycopene |
| 1 | F.carica | 0.529 | 0.163 | 0.008 | 0.029 | 0.253 | 0.002 |
| 2 | E.officinalis | 1.241 | 0.139 | 0.013 | 0.129 | 0.677 | 0.019 |
| 3 | C.indica | 0.017 | 0.387 | 0.006 | 0.013 | 0.020 | 0.007 |
| 4 | T.chebula | 0.679 | 0.545 | 0.009 | 0.088 | 0.035 | 0.011 |
| | SED | 0.0008 | 0.0015 | 0.0006 | 0.0014 | 0.0011 | 0.0012 |
| | CD(0.05) | 0.0020 | 0.01126 | 0.0013 | 0.0031 | 0.0025 | 0.0027 |

Table 1: Non Enzymatic Anti Oxidants In The Plant Samples(mg/g)

The GSH antioxidant system consist of an array of non-enzymatic and enzymatic reactions pathways involved in the neutralization of reactive free radical species. GSH is an non-enzymatic mode of defence against the free radicals³².

Green pepper fruits have vitamin C content between 52.8-115.5 mg in 100 g fresh weight^{33,34}. Plant cells contain both enzymatic and non-enzymatic antioxidants ^{35,36,37}. Ascorbic acid or vitamin C, which is the most abundant water-soluble non-enzymatic antioxidant in plants, has the ability to scavenge a wide range of ROS such as

superoxide anion, singlet oxygen and hydrogen peroxide³⁸. Ascorbate provides the first line of defense against damaging reactive oxygen species (ROS), and helps protect plant cells from many factors that induce oxidative stress, including wounding, ozone, high salinity, and pathogen attack. Ascorbate works in cooperation not only with glutathione (Halliwell-Asada cycle), but also maintains the regeneration of α - tocopherol, providing synergic protection of the membranes³⁹.

A study showed that extracts prepared from the leaves of Ficus carica L have antioxidant capacity. Antioxidant capacity results are consistent with total flavonoid and phenol contents. The α - tocopherol content of the n-hexane extract was found to be 3.2788%, whereas it was calculated as 0.0570% on the dry-weight basis of the leaves⁴⁰.

A study was carried out to evaluate the relative contribution of different polyphenols such as total phenolics, flavonoids and flavonol contents and their antioxidants activities. For this purpose the total phenolics, flavonoids and flavonol contents of some medicinal plants were determined in the aqueous extracts of leaves of *Trichosenthes dioica*, fruits of *Moringa olifera* and *Ficus bengalensis* as well as seeds of *Emblica officinalis*. Total antioxidant activity of these extracts was monitored by Free Radical Absorbing Power (FRAP) assay. They observed more phenolic content in leaves of *T.dioica* compared to the fruits and seeds of *M. oleifera* and *E.officinalis* and aerial roots of *F.bengalensis*. The highest antioxidant activity was also found in E.officinalis compared to other plant samples used in the study. These results indicated that E.officinalis seeds could be used as potential antioxidant supplement⁴¹.

The study investigated the chemistry and antioxidant properties of four commercial *E. officinalis* fruit extracts in order to determine if there are any qualitative-quantitative differences. All extracts produced positive responses in the total phenol, total flavonoid and total tannin assays. The presence of predominantly (poly)phenolic analytes, e.g. ellagic and gallic acids and corilagin, was confirmed by RP-HPLC coupled with photodiode array detection. Despite ascorbic acid being a major constituent of E. officinalis fruits, the furanolactone could not be identified in one of the samples. The extracts demonstrated varying degrees of antioxidative efficacy. The extract designated IG-3 was consistently amongst the most effective extracts in the iron(III) reduction and 1,1-diphenyl-2-picrylhydrazyl and superoxide anion radical scavenging assays while the extract designated IG-1 demonstrated the best hydroxyl radical scavenging activity. All extracts appeared to be incapable of chelating iron(II) at realistic concentrations⁴².

CONCLUSION

Plant based products have been in use for medicinal, therapeutic and other purposes right from the dawn of history. Fruits and vegetables contain significant levels of biologically active components that impart health benefits beyond basic nutrition. The study shows that *E.officinalis* contain more quantity of non enzymatic antioxidants which exhibits the benefits of consumption of the raw fruit.

REFERENCES

 Hollman P, Katan M. Absorption, metabolism and health effects of dietary flavonoids in man. Biomed Pharmacother. 1997;51:305-310.

- 2. Liu RH. Health benefits of fruit and vegetables are from additive and synergistic combinations of phytochemicals. Am J Clin Nutr. 2003;78(3Suppl):517S-520S.
- Ames B, Shigenaga M, Hagen T et al. Oxidants, antioxidants, and the degenerative diseases of aging. Proc Natl Acad Sci. 1993; 90:7915-7922
- Sirisha N, Sreenivasulu M, Sangeeta K, Madhusudhana Chetty C et al. Antioxidant Properties of Ficus Species – A Review. International Journal of PharmTech Research 2010;2(4):2174-2182
- 5. Bellakhdar J, Claisse R, Fleurentin J, Younos C et al. Repertory of standard herbal drugs in the Moraccan pharmacopoea. J Ethnopharmacol. 1991;35:123–143.
- Guarrera P.Food medicine and minor nourishment in the folk traditions of central Italy (Marche, Abruzzo and Latinum). Fitoterapia. 2003;74: 515–544
- Damodara Reddy. V, Saayi Krushna G, Padmavathi P, Varadacharyulu. N.Ch et al. Effect of *Emblica officinalis* against alcohol-induced biochemical changes in plasma and red blood cells of rats. African Journal of Biochemistry Research. 2007;1 (6): 101-105
- Wealth of India . A dictionary of Indian raw materials and India products. Raw Material, New Delhi, 1992:4: 312.
- 9. Deokate UA, Khadabadi SS. Pharmacology and phytochemistry of *Coccinia indica* Journal of Pharmacognosy and Phytotherapy. 2011;3(11):155-159,
- 10. Duraisankar M, Ravichandiran V. In vitro radical scavenging activity of *Cephalandra indica*. International Research journal of Pharmacy. 2012;3(5): 327-330
- Malekzadeh F, Ehsanifar H, Shahamat M, Levin M, Colwell RR et al. Antibacterial activity of black myrobalan (*Terminalia chebula* Retz) against *Helicobacter pylori*. International Journal of Antimicrobial Agents. 2001;18, 1, 85-88.
- 12. Aqil F, Ahmad I.Antibacterial properties of traditionally used Indian medicinal plants.Methods and Findings in Experimental and Clinical Pharmacology.2007;29(2):79-92.
- Pratibha N, Saxena VS, Amit A, D'Souza P, Bagchi M, Bagchi D et al. Anti-inflammatory activities of Aller-7, a novel polyherbal formulation for allergic rhinitis. International Journal of Tissue Reaction. 2004;26:43-51.
- Kishwar Hayat Khan. The effect of regular intake of *Terminalia chebula* on oxidative stress in mice originated from *Salmonella typhimurium*. EurAsia J BioSci . 2009;3: 113-121
- Halliwell B, Gutteridge JMC. Free radicals in biology and medicine. Third ed. Oxford: Science Publications. 2000

- Dikilitas M, Kocyigit A, Yigit F et al. A molecular-based fast method to determine the extent of DNA damages in higher plants and fungi. African Journal of Biotechnology. 2009; 8: 3118-3127.
- Thaipong K, Boonprakob U, Crosby K, Cisneros-Zevallos L, Byrne DH et al. Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruits extracts. J. Food Compos. Anal.2006;19: 669-675.
- Klimczak I, Malecka M, Szlachta M, Gliszczy_ska-_wigło A et al. Effect of storage on the content of polyphenols, vitamin C and the antioxidant activity of orange juices. J. Food Compos. Anal. 2007;20: 313-322.
- 19. Rupasinghe VHP, Clegg S et al. Total antioxidant capacity, total phenolic content, mineral elements, and histamine concentrations in wines of different fruit sources. J. Food Compos. Anal. 2007; 20: 133-137.
- Pawlowska AM, Oleszek W, Braca A et al. Quali-quantitative analyses of flavonoids of *Morus nigra* L. and *Morus alba* L. (Moraceae) fruits. J. Agric. Food Chem. 2008;56: 3377-3380.
- 21. Skerget M, Kotnik P, Hadolin M, Hraš AR, Simini M, Knez Ž et al. Phenols, proanthocyanidins, flavones and flavonols in some plant materials and their antioxidant activities. Food Chem. 2005;89: 191-198
- 22. Marinova D, Ribarova F, Atanassova M et al. Total phenolics and total flavonoids in Bulgarian fruits and vegetables. J. Univ. Chem. Technol. Metallurgy. 2005;40: 255-260
- Maisuthisakul P, Pongsawatmanit R, Gordon MH et al. Assessment of phenolic content and free-radical scavenging capacity of some Thai indigenous plants. Food Chem. 2007;100: 1409-1418.
- Jaleel CA, Manivannan P, Sankar B, Kishorekumar A, Sankari S, Panneerselvam R et al.Paclobutrazol enhances photosynthesis and ajmalicine production in *Catharanthus roseus*. Process Biochemistry. 2007a; 42: 1566–1570.
- 25. Jaleel CA, Manivannan P, Kishorekumar A, Sankar B, Gopi R, Somasundaram R, Panneerselvam R et al. Alterations in osmoregulation, antioxidant enzymes and indole alkaloid levels in *Catharanthus roseus* exposed to water deficit. Colloids and Surfaces B: Biointerfaces. 2007b;59:150–157.
- 26. Roe JH, Heather CA. The determination of Ascorbic acid in whole blood and urine through 2,4 Dinitro phenyl hydrazine derivative of dehydro Ascorbic acid.J.Biol.Chem.1953;147:399-407.

- 27. Rosenberg HR.Chemistry and physiology of the vitamins. Interscience Publishers, Inc. NewYork. 1992;452-453.
- 28. Moron MS, Depierre JW, Mannervik B et al.Levels of Glutathione, Glutathione Reductase and Glutathione transferase activities in rat lung and liver.Biochem.Biophya Acta.1979;582:67-68.
- 29. Malick, CP, Singh MB.Plant enzymology and istoenzymology.Kalyani Publications, NewDelhi. 1980;286.
- 30. Zakaria H, Sinpson K, Brown PR, Krotatin A et al. Use of reversed phase high performance liquid chromatographic analysis for the determination pro vitamin A in tomatoes.J.Chromatography.1979;176:109-117.
- 31. Zakaria H, Sinpson K, Brown PR, Krotatin A et al.Use of reversed phase high performance liquid chromatographic analysis for the determination pro vitamin A in tomatoes. J.Chromatography.1979;176:109-117
- Malarvannan L, Devaki T. Protective effect of indigofra tinctoria on tissue anti oxidant damage system against D-Galactose amine and Endotoxin induced Hepatopathy in rats. J.of Natural remedies. 2003;31:49-53.
- 33. Howard R, Talcott ST, Brenes CH, Villalon B et al.Changes in phytochemical and antioxidant activity of selected pepper cultivars (Capsicum species) as influenced by maturity. Journal of Agricultural and Food Chemistry 2000;48: 1713-1720.
- 34. Suntomsuk L, Gritsanpun W, Nilkamhank S, Paochom A et al. Quantitaiton of vitamin C content in herbal juice using direct titration. Journal of Pharmaceutical and Biomedical Analysis. 2002; 28: 849-855.
- 35. Mittler R, Feng XQ, Cohen M et al. Posttranscriptional suppression of cytosolic ascorbate peroxidase expression during pathogen induced programmed cell death in Biochemistry. 1998;48: 337-350.
- 36. Riedle-Bauer M. Role of reactive oxygen species and antioxidant enzymes in systemic virus infections of plants. Journal of Phytopathology. 2000;148: 297-302
- 37. Kaya C, Tuna AL, Dikilitas M, Cullu MA et al. Responses of some enzymes and key growth parameters of salt-stressed maize plants to foliar and seed applications of kinetin and indole acetic acid. Journal of Plant Nutrition. 2010;33: 405-422.
- 38. Suza WP, Avila CA, Carruthers K, Kulkarni S, Goggin FL, Lorence A et al.Exploring the impact of wounding and jasmonates on ascorbate metabolism. Plant Physiology and Biochemistry.2010;48:337-350

- 39. Thomas CE, McLean LR, Parker RA, Ohlweiler DF et al. Ascorbate and phenolic antioxidant interactions in prevention of liposomal oxidation. Lipids.1992;27: 543-550.
- 40. Sibel Konyaloglu, Husniye Saglam, Bijen Kvcak et al. Tocopherol, Flavonoid, and Phenol Contents and Antioxidant Activity of Ficus carica Leaves Pharmaceutical Biology. 2005a ; 43(8): 683–686
- 41. Sharma RK, Sanjukta Chatterji, Devendra K Rai, Shikha Mehta, Prashant K Rai, Rakesh K Singh, Geeta Watal, Bechan Sharma et al. Antioxidant activities and phenolic contents of the aqueous extracts of some Indian medicinal plants. Journal of Medicinal Plants Research 2009; 3(11):944-948
- 42. Poltanov EA, Shikov AN, Dorman HJ, Pozharitskava ON, Makarov VG, Tikhonov VP, Hiltuman R et al. Chemical and antioxidant evaluation of Indian gooseberry *(Emblica officinalis Gaertn., syn. Phyllanthus emblica L.)* supplements.Phytother res. .2009; 23(9):1309-15.