

Research article

International Journal of Research in Pharmacy and Science

Alteration in Thyroid Hormones and Vitamins As Early Markers of Aluminum Induced Neurodegeneration in Rats

Pal Amit¹, Choudhary Manisha², Joshi Devesh Kumar², Tripathi Sandeep^{2*}, Modi Dinesh Raj¹

¹Department of Biotechnology, Babasaheb Bhimrao Ambedakar University, Lucknow (UP), India ²Department of Biotechnology, NIET, NIMS University, Shobha Nagar, Jaipur-303121, India

ABSTRACT

Aluminum (Al) is a potent neurotoxic element, which has been suggested to play an important role in the degeneration of nerve cells of experimental animals as well as human brain. It is associated with Alzheimer's like symptoms leading to cognitive decline. In the present study we attempt to investigate the early markers of aluminum induced neurodegeneration in rats. The effects of Al on thyroid function and vitamins were evaluated in adult wistar rats and its correlates with the Al induced cognitive dysfunction. 100 mg /kg body weight of AlCl₃ was given orally to rats for 90 days. Time dependent study was carried out on the 30, 60 and 90 days of treatment. The T3, T4, FT4 and Vitamins A, C, E was estimated on 30, 60 and 90 days along with behavioral test. We observed that serum concentration of total T3 and T4 was decreased and FT4 was insignificant changed while vitamins C and E were markedly changed on 60 and 90 days of treatment. It has been observed that the concentration of T3 and T4 levels and vitamin E and C is significantly correlates with the behavioral activity. On the basis of results it may conclude that the concentration of T3, T4, Vitamin E and Vitamin C may be used as a marker of cognitive decline for the detection of aluminum induced neurobehavioral changes.

KEYWORDS: Thyroid hormone; Vitamins; Aluminum; Neurodegeneration; Cognitive impairment

Corresponding Author Dr. Sandeep Tripathi Assistant Professor, Department of Biotechnology NIET, NIMS University, Shobha Nagar Jaipur-303121 India E-mail: <u>sandeeptripathiphd@gmail.com</u>

INTRODUCTION

Aluminum (Al) is a third most abundant element in the earth's crust and used extensively in modern daily life. It is found in our food products, medicines and also added to drinking water for purification purposes. The high Al diet may lead to increase the deposition of Al in the CNS¹. It is known to be a potential contributing factor in the etiology of severe neurodegenerative disorders such as Alzheimer's disease $(AD)^{2,3}$. Al promotes the formation of amyloid- β protein plaques^{4,5} by aggregating tau proteins in Alzheimer's disease⁶. Our previous studies^{1, 7,8} reported that Al increases oxidative burden in the brain and alters post synaptic density and cognitive impairment in the animals⁹. Moreover, Bolla and collogues¹⁰ also observed that decline in visual memory in hemodialyzed patients who exhibited higher serum Al.

In addition, It is reported that plasma triiodothyronine (T3) and thyroxine (T4) concentrations were increased in sublethally Al-stressed brown trout (Salmo trutta)¹¹. Experimental studies reported that changes in thyroid hormones induce deposition of amyloid- β , the major component of the amyloid deposits found in the brain of cases of AD. Thyroid hormone plays a significant role in adult brain function, but the precise mechanisms still unknown^{12,13}.

Vitamins in the body act as potent antioxidant. Vitamin like A, C, E and beta Carotene is a naturally occurring antioxidant nutrient that plays an important role in animal health by inactivating harmful free radicals that are produced during normal metabolic process or induced by external intoxicants such as Al. Vitamin (ascorbic acid) is an essential micronutrient required for normal metabolic functioning of the body and it ameliorates glial activation and reduces release of proinflammatory cytokines induced by Al^{14} . Previously, it is known that antioxidant nutrients, like vit-C, are important for neurological function¹⁵⁻¹⁷. Retinoic acid (RA), the active metabolite of vitamin A (retinoid), has also been shown to control the expression of genes related to APP processing^{18,19} in AD. Deprivation of vitamin A results in Aβ accumulation²⁰, loss of hippocampal long-term potentiation (LTP)²¹ and memory deficits in rodents^{22,23}, all of which are hallmarks of AD. The impairment in spatial learning and memory and the depression of synaptic plasticity that occurs in vitamin A-deprived rodents also occur during aging in rodents²⁴.

Taking into consideration the above facts, the present study was conducted with an objective to evaluate the thyroid status, serum vitamin levels and behavioural activity in AL toxicity and its correlate them with the time dependent severity of the disease process.

MATERIAL AND METHODS

ANIMALS

Albino Wistar rats (weight 220 ± 10 g, n = 20) were purchased from Industrial Toxicology Research Institute, Lucknow, (UP) India. The animals were separately housed in polypropylene cages in a room, which was maintained at a temperature of 22 \pm 2 ⁰C, relative humidity of 50 \pm 10 % and 12h light dark cycles. They were fed a commercial pellet diet (Dayal Industries, Barabanki, UP, India) and allowed access to water ad libitum. The Institutional Animal Ethics Committee approved the study prior to the initiation of the experiment and also approved all experimental protocols.

TREATMENT

Animals were randomly divided into two groups (n = 10) viz. control and experimental. Aluminum chloride 100 mg/ kg body weight mixed with 1% gum acacia formed an aqueous suspension which was directly introduced into the rat pharynx via a feeding cannula (The sharp age of the tip of a hypodermic needle no. 16 was blunted by grinding on a stone and thereafter bent to 120° so that the curved needle could easily be introduced into the rat pharynx via oral cavity without the pointed tip lacerating the passage) to experimental group and an equivalent volume of physiological saline was given to control groups for 90 days.

BEHAVIORAL TEST

- 1. Spontaneous motor activity [SMA]: SMA was measured by scored on a scale of 0 9 in which SMA in control group was assigned score 4.
- 2. Muscle coordination test (Rota rod): The period of stay on rotating rod (speed: 5 rotations / min; Total duration of test 2 min) for each control and treated rat were recorded by Rotamex (Techno Electronics, India). The rats were trained to stay for period of 2 min on rotating rod and only trained rats were included in the study. Motor was measured using Rota Rod at least 5s and it was rotated at speed of 10 rpm for 2 consecutive days on third day the time duration of each rotation speed was also recorded.

3. Y-maze learning and memory test

A test was used to assess the short term memory 'in vivo' was carried out in both Al treated and normal rats by Y maze (techno co. 40 cm long x 13 cm height x 10 cm width). Y maze test is a gross test for spatial memory. This test was used to see if the mouse remembers the arm it had just explored and therefore enters in one of the other arms of the maze. Rats were placed at the bottom of the Y (middle arm) in the y maze and are allowed to explore freely all three arms for an eightminute session. The first two minutes are for habituation and for the last six minutes the alteration between arms was recorded via photo beam breaks. The acquisition time was noted to determine the short-term memory.

BLOOD COLLECTION

After 30, 60 and 90 days of treatment the blood was collected from the tail vein of the rats in heparinized vial and in plain vial for hemolysate preparation and for serum separation respectively.

HEMATOLOGY

The hematological tests were carried out using commercially available Qualigens kit. The hematological parameters namely hemoglobin (Hb; mg/dl), red blood corpuscles (RBC; X 10^6 cells/ mm³ of blood), white blood corpuscles (WBC; X 10^3 cells/ mm³ of blood), packed cell volume (PVC; percent), prothrombin time (PTT; sec) and erythrocyte sedimentation rate (ESR; mm/h) were carried out in blood samples of Al treated young and old rats and their controls.

SERUM ENZYME LEVELS

Serum enzymes aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), urea, creatine and glucose were assayed using commercially available Qualigens kits (Mumbai, India).

HORMONAL ASSAY

Serum T3 (triiodothyronine), T4 (thyroxine), and FT4 (free tetraiodothyronine) were analysed using commercially available kit (DPC-USA) through solid phase radioimmuno assay.

VITAMIN ESTIMATIONS IN SERUM

Vitamin A was estimated by the previously described procedure of Craft et al^{25} using high pressure liquid chromatography (HPLC) system (Waters Limited Mulford USA). The retinyl acetate was used as a internal standard and retinol levels were expressed as $\mu g/dl$. Serum vit-C was measured according to method of Lowry et al^{26} using 2,4 dinitrophenyl hydrazine in presence of thiourea as a mild reducing reagent to form a red coloured compound bis-hydrazone, which was measured at 520nm in spectrophotometer. Serum vitamin-E was estimated by method of Baker & Frank after xylene

extraction & reduction of ferric to ferrous ions, which then forms a red coloured complex with α - α ' Dipyridyl²⁷. Absorbance was read at 460nm by spectrophotometer and a correction for the carotenes was made after adding ferric chloride and measured at 520 nm.

RESULT AND DISCUSSION

In the present study we observed that Al induced neurobehavioral changes in rats and correlates with the time dependent association of haematological, hormonal and biochemical findings. The effect of Al on body weight gain, food intake and feed efficiency was progressively increased during the experimental period of both the groups. The final body weight of intoxicated rats (Fig.1) with Al was significantly lower than that of the health normal group. These results clearly indicate that Al cause a significant decrease in body weight. This harmful effect of Al on the body weight gain was elevated paralleled with increase of Al dose. The amount of food intake of both groups was unchanged significantly. This means that the value of food intake was not paralleled to the rate of growth and feed efficiency. The obtained results are in agreement with the findings of our previous study⁷. It is suggestive that loss of body mass indicated that Al induces severe toxicity and it may be due to loss of lipid, protein and other biomolecules.

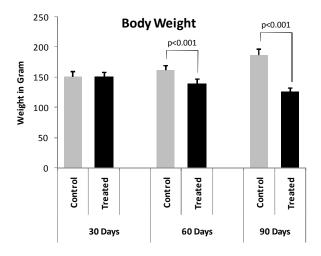


Fig.1. Time dependent effect of Al administration on the body weight of rats.

Values are expressed as mean \pm SEM for ten animals (N=10) in each group. Statistical significance was determined by Mann-Whitney *p*-test. Probability, *p*-value less than 0.05 were considered statistically significant of control and treated groups on different intervals.

Earlier Studies in animal models shows the loss of cognitive function under Al toxicity. The behavioral profiles (Fig.2) exhibited spontaneous motor activity (SMA) was found to be significantly

increased in Al treated rats as compared with the controls on the day 60 and 90 (Fig-2a). On the other hand muscle in-cordination was found to be increased at 90 days (Fig. 2b). The Y maze learning and memory test showed (Fig. 2c) that there was gradually decreased in acquisition time in Al treated rats as comparison with control rats. These results were significantly appeared from the day 60.

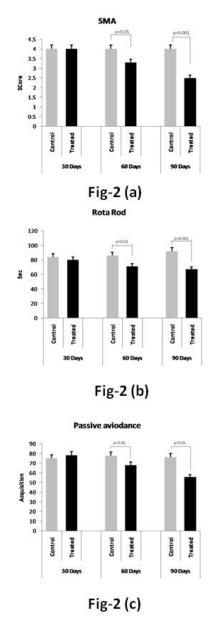


Fig. 2: Time dependent effect of Al administration on behavioural activity (spontaneous motor activity, rota rod and passive avoidance).

Values are expressed as mean \pm SEM for ten animals (N=10) in each group. Statistical significance was determined by Mann-Whitney *p*-test. Probability, *p*-value less than 0.05 were considered statistically significant of control and treated groups on different intervals.

Parameters	30 Days		60 1	Days	90 days	
	Control	Al treated	Control	Al treated	Control	Al treated
Hb	12.8 ± 0.2	12.1 ± 0.1	12.9 ± 0.13	$10.3 \pm 0.12*$	11.7 ± 0.34	9.5 ± 0.4*
RBC	4.54 ± 0.1	4.22 ± 0.1	4.71 ± 0.06	3.41 ± 0.08	3.94 ± 0.14	$2.66 \pm 0.1*$
WBC	6.43 ±0.1	7.32 ± 0.1	6.12 ±0.11	8.02 ± 0.12	6.71 ±0.14	8.34 ±0.1*
PCV	64.8 ±2.1	65.2 ± 2.2	63.6±3.12	55.6 ± 2.12*	56.2 ±2.18	42.3 ±2.6*
РТТ	14.8 ±0.1	15.1 ± 0.2	14.3 ±0.15	13.2 ± 0.23	13.4 ±0.09	20.4 ±2.1*
ESR	4.12 ±0.1	5.1 ± 0.1	4.4 ±0.12	7.8 ± 0.14*	5.67 ±0.1	10.27 ±1.1*

Table-1. Time dependent effect of aluminum on haematological parameters

Values are expressed as mean \pm SEM for ten animals (N=10) in each group. Haemoglobin (Hb; mg/dl), red blood corpuscles (RBC; X 10⁶ cells/ mm³ of blood), white blood corpuscles (WBC; X 10³ cells/ mm³ of blood), packed cell volume (PVC; percent), prothrombin time (PTT; sec) and erythrocyte sedimentation rate (ESR; mm/h) concentration on 30, 60 and 90 day of treatment of aluminum. Statistical significance was determined by Mann-Whitney *p*-test. Probability, (*p*< 0.05) was considered statistically significant control and treated groups on different intervals.

The haematological modifications induced by Al demonstrate that altered peripheral blood composition (Table-1) is a reflection of disrupted haematopoietic process. Blood, which rapidly and constantly flows through the tissues and play in important role in the transportation of nutrients, antioxidants, hormones and some other chemicals. These chemical are required to for the physiological functioning of the body. Al supplementation caused decrease in RBCs and WBCs count and it is time dependent effect of Al intoxication. After long term exposure it is found that detritus alteration was observed these finding concomitant with the observation of Mahieu et al²⁸ and Zaman et al²⁹. Moreover, the haemoglobin concentration and PVC after Al treatment were also found to be gradually decreased with the increasing of exposure time. There is evidence that anaemia is associated with Al accumulation in plasma and/or bone tissue in patients with chronic renal insufficiency³⁰. The present results indicated that aluminium treatment resulted in a significant (P < 0.05) decline in haemoglobin (Hb), total erythrocytic count (TEC) and packed cell volume (PCV), while total leukocyte count (WBC) increased. Vittori et al³⁰ reported that erythrocyte morphological changes were induced by aluminium and cells lost their typical biconcave shape. Simultaneously, an increased membrane protein breakdown compatible with band 3 degradation was detected. They also suggested that aluminium may disturb erythropoiesis through combined effects on mature erythrocytes and

cellular metabolism in late erythroid progenitors. Also, the inhibition in erythropoiesis and iron metabolism due to aluminium treatment probably hinders haemoglobin synthesis and erythroid Humans are uniformly exposed to Al that is present in the soil, food and drinking water. There are several data linking elevated Al levels to neurological pathologies such as multiple sclerosis, Guam Parkinson dementia, Parkinson's disease and Alzheimer's disease³¹. It was reported that prolonged Al sulphate intake accelerate features of senescence in the adult mice liver. In our study we found that the elevated concentration of serum ALP, GOT and GPT followed by Al administration on days 30, 60 and 90 (Table 2).

Test	30 days		60 1	Days	90 days	
	Control	Treated	Control	Treated	Control	Treated
ALP (KA)	25.6 ±1.0	46.3 ±4.2*	27.9 ±2.21	77.7 ±3.3*	26.3 ±3.4	85.81±3.5*
SGOT(U/ml)	17.3 ±1.8	20.3 ±1.3*	16.4 ±1.64	28.2 ±2.1*	17.9 ±1.3	23.2 ±1.7*
SGPT(U/ml)	13.2 ±1.2	12.4 ±1.9	11.9 ±1.65	16.4 ±1.91*	12.9 ±1.37	19.2 ±1.1*

Table-2. Time dependent effect of aluminum on clinical enzymes

Values are expressed as mean \pm SEM for ten animals (N=10) in each group. Alkaline phosphate (ALP; KA), serum oxaloacetic acid transaminase (SGOT; U/ml) and serum glutamine pyruvate transaminase (SGPT; U/ml) activity on 30, 60 and 90 day of aluminum treatment. Statistical significance was determined by Mann-Whitney *p*-test. Probability, *p*-value less than 0.05 were considered statistically significant control and treated groups on different intervals.

The Al toxicity was characterized by markedly elevated serum ALP, GOT and GPT on days 60 and 90 as compared with their respective controls. Transaminase i.e. SGOT and SGPT are excellent markers of tissue damage. Elevated in serum level has also been reported in the Al induced neurodegeneration⁸. On the other hand, investigation of the subcellular distribution of Al in postmortem examination of the dialysis dementia patient indicates deposition of Al in brain and liver.

Prolonged Al exposure accelerates ageing changes in the adult rat brain³² and enhanced Al deposition in the brain is a shared characteristic of progressive neurological diseases that are common in aged populations³³. The deposition of Al in non nervous organs and its subsequent effects are less known. It was previously described that the effects of Al in the rat kidney and liver, where it induces lysosomal activation, increases iron deposition³⁴ and hence the liver was involved in Al absorption and excretion through biliary flux³⁵.

In this study, the thyroid hormone (i.e. T3, T4 and FT4) were assessed and found that significantly altered concentration of T3 and T4 levels. However, FT4 were found to be insignificant change in different intervals (Table.3). Both T3 and T4, changes were observed on day 60 and 90 of Al treatment

Tripathi Sandeep et al. IJRPS 2012,2(2),137-150

when compared with their respective controls. In healthy individuals the relationship between T3 and T4 is finely controlled in such a way that differences in hormone activity due to the wide range of serum T4 among normal's may be buffered by systematic adjustment of the T3/T4 ratio. Deficiency of thyroid hormone during environmental exposure of brain is associated with profound, and often irreversible morphological defects that contribute to severe cognitive and neurological impairment³⁶. Although the developmental effects of thyroid hormone have been well established, its influence on the adult brain is relatively poorly understood. The adult mammalian brain does not exhibit the severe morphological defects associated with developmental hypothyroidism, however, hypothyroidism in adulthood has been clearly linked to cognitive dysfunction and depressed mood^{37,38}. Neurophysiological stress has been reported to decrease plasma levels of vitamins A, E, and Vit C^{39,40}. During the past several years, major epidemiological studies have addressed the role of the antioxidant vitamins A, C, and E in the protection against different diseases.

Test	30 days		60 c	lays	90 days		
	Control	Treated	Control	Treated	Control	Treated	
T3	1.86 ± 0.22	2.02 ± 0.1	2.06 ± 0.22	3.12±0.1*	2.33 ± 0.19	4.3 ± 0.23*	
T4	68.7 ± 1.2	73.5 ± 2.0	62.5 ± 1.2	42.4 ±2.9*	69.8 ± 1.3	41.2 ±1.7*	
FT4	11.5 ±1.2	10.03 ± 1.6	11.50 ± 1.2	12.03±1.6	17.24 ± 2.08	15.93 ±1.6	

Values are expressed as mean \pm SEM for ten animals (N=10) in each group. Alkaline phosphate (ALP; KA), serum oxaloacetic acid transaminase (SGOT; U/ml) and serum glutamine pyruvate transaminase (SGPT; U/ml) activity on 30, 60 and 90 day of aluminum treatment. Statistical significance was determined by Mann-Whitney *p*-test. Probability, *p*-value less than 0.05 were considered statistically significant control and treated groups on different intervals.

Al caused a significant decrease in the level of vitamins in blood serum of rats. Vitamin C concentration was diminished on the 60^{th} and 90^{th} days. Administration of Al caused a significant decrease in vitamin A levels in the blood serum on the 60^{th} and 90^{th} days. Al, similar to the case of other vitamins, caused a significant decrease in vitamin E on the 30^{th} , 60^{th} and 90^{th} days (Table.4).

Vitamin E (VE) (alpha-tocopherol) is a naturally occurring antioxidant nutrient that plays an important role in animal health by inactivating harmful free radicals that are produced through normal cellular activity and from various stressors. Fatma and El-Demerdash⁴¹ reported that elevated Al toxicity can be prevents by the supplementation of vitamin E

Tripathi Sandeep et al. IJRPS 2012,2(2),137-150

Test	30 days		60 days		90 days	
	Control	Treated	Control	Treated	Control	Treated
Vit A (umole/l)	1.32 ± 0.03	$1.1\ 2\pm 0.02$	1.41 ± 0.03	1.22 ±0.02*	1.27 ± 0.03	$0.74 \pm 0.01*$
Vit C (umole/l)	1.09 ± 0.09	0.96 ± 0.08	1.17 ± 0.07	0.85 ±0.08*	1.12 ± 0.07	0.79 ±0.07*
Vit –E (umole/l)	30.9 ± 0.1	23.9 ±0.1*	33.6 ± 0.3	18.5 ±0.2*	31.2 ± 0.3	$16.6 \pm 0.2*$

Table-4. Time dependent effect of aluminum on vitamins level (Vit A, VitC and Vit E) levels.

Values are expressed as mean \pm SEM for ten animals (N=10) in each group. Vitamin A, C and E on 30, 60 and 90 day of aluminum treatment. Statistical significance was determined by Mann-Whitney *p*-test. Probability, *p*-value less than 0.05 were considered statistically significant control and treated groups on different intervals.

On the other hand, Yousef⁴² has shown the protective effect of Vitamin C on Al induced changes in haemato-biochemical parameters. The results of the present study, the reduced concentration of vitamin particularly, Vit E and C may be responsible for the excessive generation of free radical in the brain. The reduced form of the vitamin, ascorbic acid, is an especially effective antioxidant owing to its high electron-donating power and ready conversion back to the active reduced form. There is great interest in the clinical roles of Vitamin C because of evidence that oxidative damage is a root cause of several neurodegenerative disorders. Population studies show that individuals with high intakes of Vitamin C have lower risk of a number of chronic diseases, including heart disease, cancer, eye diseases, and neurodegenerative conditions.

CONCLUSION

On the basis of results it may be safely conclude that Al induced changes in haematological parameters, clinical enzymes, thyroid hormones and vitamins are correlates with the neurodegenerative disorders as evident by the neurobehavioral changes following 90 days of Al treatment. Time dependent changes may be used as the markers of neurodegenration. Further study is also required to get depth knowledge relating to assess aluminium induced neuronal changes non- invasively.

REFERENCES

 Tripathi S, Mahdi AA, Nawab A, Chander R, Hasan M, Siddiqui MS, Mahdi F, Mitra K, Bajpai VK. Influence of age on aluminum induced lipid peroxidation and neurolipofuscin in frontal cortex of rat brain: a behavioral, biochemical and ultrastructural study. Brain Res. 2009; 1253: 107-116.

- McLachlan DR, Lukiw WJ, Kruck TP. New evidence for an active role of aluminum in Alzheimer's disease. Can J Neurol Sci. 1989; 16(4): 490-497.
- Exley C and Esiri MM. Severe cerebral congophilic angiopathy coincident with increased brain aluminum in a resident of Camelford, Cornwall, UK. J. Neurol. Neurosurg. Psychiatry. 2006; 77(7): 877-879.
- Jiang T, Zhi XL, Zhang YH, Pan LF, Zhou P. Inhibitory effect of curcumin on the Al(III)induced Aβ (42) aggregation and neurotoxicity in vitro. Biochim Biophys Acta. 2012; 1822(8): 1207-1215.
- Kawahara M and Kato-Negishi M. Link between Aluminum and the Pathogenesis of Alzheimer's Disease: The Integration of the Aluminum and Amyloid Cascade Hypotheses. Int J Alzheimers Dis. 2011; 2011: 276393.
- Savory J, Huang Y, Wills MR, Herman MM. Reversal by desferrioxamine of tau protein aggregates following two days of treatment in aluminum-induced neurofibrillary degeneration in rabbit: implications for clinical trials in Alzheimer's disease. Neurotoxicology. 1998; 19(2): 209-214
- Tripathi S, Mahdi AA, Hasan M, Mitra K, Mahdi F. Protective potential of Bacopa monniera (Brahmi) extract on aluminum induced cerebellar toxicity and associated neuromuscular status in aged rats. Cell Mol Biol. 2011; 57(1): 3-15
- Tripathi S, Somashekar BS, Mahdi AA, Gupta A, Mahdi F, Hasan M, Roy R, Khetrapal CL. Aluminum mediated metabolic changes in rat serum and urine: A proton magnetic resonance study. J Bio Mol Toxicol. 2008; 22(2): 119-127.
- Thirunavukkarasu SV, Venkataraman S, Raja S, Upadhyay L. Neuroprotective effect of Manasamitra vatakam against aluminium induced cognitive impairment and oxidative damage in the cortex and hippocampus of rat brain. Drug Chem Toxicol. 2012; 35(1): 104-115.
- 10. Bolla KI, Briefel G, Spector D, Schwartz BS, Wieler L, Herron J, Gimenez L. Neurocognitive effects of aluminum. Arch Neurol. 1992; 49(10): 1021-1026.
- Waring CP, Brown JA, Collins JE, Prunet P. Plasma prolactin, cortisol, and thyroid responses of the brown trout (Salmo trutta) exposed to lethal and sublethal aluminium in acidic soft waters. Gen Comp Endocrinol. 1996; 102(3): 377-385.

- Bauer RM, Stadler TC, Bader MJ, Bastian PJ, Gozzi C, Stief CG. 25 years of progress in urology: breakthrough for sexual dysfunction, testicular cancer and incontinence. MMW Fortschr Med. 2008; 150(48): 144-145.
- 13. Davis JD and Tremont G. Neuropsychiatric aspects of hypothyroidism and treatment reversibility. Minerva Endocrinol. 2007; 32: 49-65.
- 14. Nedzvetsky VS, Tuzcu M, Yasar A, Tikhomirov AA, Baydas G. Effects of vitamin E against aluminum neurotoxicity in rats. Biochemistry (Mosc). 2006; 71(3): 239-244.
- Daskalopoulos R, Korcok J, Tao L, Wilson JX. Accumulation of intracellular ascorbate from dehydroascorbic acid by astrocytes is decreased after oxidative stress and restored by propofol. Glia. 2002; 39(2): 124-132.
- Rice ME. Ascorbate regulation and its neuroprotective role in the brain. Trends Neurosci. 2000; 23(5): 209-216.
- 17. Siushansian R and Wilson JX. Ascorbate transport and intracellular concentration in cerebral astrocytes. J Neurochem. 1995; 65(1): 41-49.
- 18. Culvenor JG, Evin G, Cooney MA, Wardan H, Sharples RA, Maher F, Reed G, Diehlmann A, Weidemann A, Beyreuther K, Masters CL. Presenilin 2 expression in neuronal cells: induction during differentiation of embryonic carcinoma cells. Exp Cell Res. 2000; 255(2): 192-206.
- 19. Satoh J and Kuroda Y. Beta-catenin expression in human neural cell lines following exposure to cytokines and growth factors. Neuropathology. 2000; 20(2): 113-123.
- 20. Corcoran JP, So PL, Maden M. Disruption of the retinoid signaling pathway causes a deposition of amyloid beta in the adult rat brain. Eur J Neurosci. 2004; 20(4): 896-902.
- 21. Misner DL, Jacobs S, Shimizu Y, de Urquiza AM, Solomin L, Perlmann T, De Luca LM, Stevens CF, Evans RM. Vitamin A deprivation results in reversible loss of hippocampal longterm synaptic plasticity. Proc Natl Acad Sci U S A. 2001; 98(20): 11714-11719.
- 22. Cocco S, Diaz G, Stancampiano R, Diana A, Carta M, Curreli R, Sarais L, Fadda F. Vitamin A deficiency produces spatial learning and memory impairment in rats. Neuroscience. 2002; 115(2): 475-82.
- 23. Etchamendy N, Desmedt A, Cortes-Torrea C, Marighetto A, Jaffard R. Hippocampal lesions and discrimination performance of mice in the radial maze: sparing or impairment depending on the representational demands of the task. Hippocampus.2003; 13(2): 197-211.

- 24. Etchamendy N, Enderlin V, Marighetto A, Vouimba RM, Pallet V, Jaffard R, Higueret P. Alleviation of a selective age-related relational memory deficit in mice by pharmacologically induced normalization of brain retinoid signaling. J Neurosci. 2001; 21(16): 6423-6429.
- 25. Craft NE, Bulux J, Valdez C, Li Y, Solomons NW. Retinol concentrations in capillary dried blood spots from healthy volunteers: method validation. Am J Clin Nutr. 2000; 72(2): 450-454.
- 26. Lowry OH, Lopez JA, Bessey OA. The determination of ascorbic acid in small amounts of blood serum. J Biol Chem. 1945; 160: 609-615.
- Baker H and Frank O. Vitamin status in metabolic upsets. World Rev Nutr Diet. 1968; 9: 124-160.
- 28. Mahieu S, del Carmen Contini M, Gonzalez M, Millen N, Elias MM. Aluminum toxicity. Hematological effects. Toxicol Lett. 2000; 111(3): 235-242.
- Zaman K, Zaman A, Batcabe J. Hematological effects of aluminum on living organisms. Comp Biochem Physiol C. 1993; 106(2): 285-293.
- Vittori D, Garbossa G, Lafourcade C, Pérez G, Nesse A. Human erythroid cells are affected by aluminium. Alteration of membrane band 3 protein. Biochim Biophys Acta. 2002; 1558 (2): 142-150.
- 31. Nayak, P. Aluminum: impacts and disease. Environ. Res. 2002; 89(2): 111-115.
- 32. Deloncle R, Huguet F, Fernandez B, Quellard N, Babin P, Guillard O. Ultrastructural study of rat hippocampus after chronic administration of aluminuml-glutamate:an acceleration of the aging process. Exp. Gerontol. 2001; 36(2): 231–244.
- 33. Miu AC, Olteanu A, Miclea M. A behavioural and ultrastructural dissection of the interference of aluminium with aging. J Alzheimers Dis. 2004; 6(3): 315-328.
- 34. Stacchiotti A, Rodella LF, Ricci F, Rezzani R, Lavazza A, Bianchi R. Stress proteins expression in rat kidney and liver chronically exposed to aluminium sulphate. Histol. Histopathol. 2006; 21(2): 131–140.
- 35. Gonzales M, Alvarez Mdel L, Pisani G, Bernal C, Roma M, Carrillo M. Involvement of oxidative stress in the impairment in biliary secretory function induced by intraperitoneal administration of aluminium to rats. Biol Trace Elem Res. 2007; 116(3): 329-348.
- 36. Porterfield SP and Hendrich CE. The role of thyroid hormones in prenatal and neonatal neurological development-current perspectives. Endocr. Rev. 1993; 14(1): 94–106.
- Dugbartey AT. Neurocognitive aspects of hypothyroidism. Arch. Intern. Med. 1998; 158(13): 1413–1418.

- 38. Jackson IM. The thyroid axis and depression. Thyroid. 1998; 8(10): 951–956.
- 39. Betti M, Minelli A, Ambrogini P, Ciuffoli S, Viola V, Galli F, Canonico B, Lattanzi D, Colombo E, Sestili P, Cuppini R. Dietary supplementation with α-tocopherol reduces neuroinflammation and neuronal degeneration in the rat brain after kainic acid-induced status epilepticus. Free Radic Res. 2011; 45(10): 1136-1142.
- 40. Lagowska-Lenard M, Stelmasiak Z, Bartosik-Psujek H. Influence of vitamin C on markers of oxidative stress in the earliest period of ischemic stroke. Pharmacol Rep. 2010; 62(4): 751-756.
- 41. Fatma M El-Demerdash. Antioxidant effect of vitamin E and selenium on lipid peroxidation, enzyme activities and biochemical parameters in rats exposed to aluminium J Trace Elem Med Biol. 2004; 18(1): 113-121.
- Yousef MI. Aluminium-induced changes in hemato-biochemical parameters, lipid peroxidation and enzyme activities of male rabbits: protective role of ascorbic acid Toxicology. 2004; 199(1): 47–57.